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Biomimetic Cleavage of RNA Models Promoted by a Dinuclear Zn(II) Complex in Ethanol. Greater than 30 kcal/mol Stabilization of the Transition State for Cleavage of a Phosphate Diester

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Abstract: The cleavage of a series of seven substituted anyl 2-hydroxypropyl phosphates (1a-g) promoted by a dinuclear Zn(II) complex $(3:Zn(II)_2:(^{-}OCH_2CH_3))$ was investigated in ethanol at \S pH 9.0 \pm 0.2 and 25 °C. The kinetics for appearance of the product phenols follow very strong saturation behavior for all substrates where the dissociation constant of the bound complex has an upper limit of $K_m = 3 \times 10^{-7}$ M and the $k_{cat}^{max \ corr}$ values (corrected for triflate inhibition) range from 168 to 3 s⁻¹. A partial $\$pH/\log k_{cat}^{max \ corr}$ profile for the **3**:Zn(II)₂:($^{-}OCH_2CH_3$)-catalyzed reaction of 1e (3-methoxyphenyl 2-hydroxypropyl phosphate) is bell-shaped, plateauing from 7.9–10, and is fit to a two kinetically important ionizations having $\$pK_a$ values of 7.22 and 10.9. The Brønsted plot of log ($k_{cat}^{max \ corr}$) vs the $\$pK_a$ values for the phenols shows a break at about 14.3 with two β_{lg} values of -1.12 and 0.0. This is analyzed in terms of a change in rate limiting step from cleavage of the phosphate to a conformational change where the binding of the phosphate changes from one P–O⁻---Zn(II) interaction to a Zn(II)---O–P-O---Zn(II) double activation. An energetics calculation comparing the ethoxide promoted cleavage of 1a-g with the $3:Zn(II)_2:(^{-}OEt)$ promoted reaction indicates that the complex, $3:Zn(II)_2$, stabilizes the ethoxide plus substrate transition state for the cleavage of 1a-g by between 33 and 36 kcal/mol. The origins of the large stabilization are discussed in terms of the effect of the medium on the various rate and equilibrium constants involved.

1. Introduction

The stability of phosphate diesters toward solvolytic cleavage is vitally tied to the biological roles of DNA and RNA as guardians of genetic information in all living systems. In the absence of a catalyst or an enzyme, phosphate diesters are extremely stable toward solvolytic cleavage but in the presence of phosphodiesterase enzymes rate accelerations of 10^{15-17} -fold¹ are achieved. Many of these enzymes contain metals ions such as Zn²⁺, Ca²⁺, Mg²⁺, Fe³⁺, and Mn²⁺ and the efficiency of such

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metalloenzymes has, not surprisingly, sparked much effort in constructing simpler catalytic systems for the cleavage of phosphate esters.^{2,3}

Recent work from this laboratory^{4,5} has focused on the catalytic cleavage of a series of aryl 2-hydroxylpropyl phosphates (1a-g; simplified models of RNA) and aryl methyl phosphates (2a-n; models of DNA) promoted by the dinuclear Zn(II) and Cu(II) complexes of 1,3-bis- N_1 , N_1' -(1,5,9-triazacy-clododecyl)propane (3) in methanol. Detailed mechanistic investigation led to the proposal that the catalyzed cleavage follows a multistep pathway consistent with a minimal process given in Scheme 1 consisting of a bimolecular binding step of the catalyst to the phosphate followed by a rearrangement to



form the catalytically active species. This is followed by one or more chemical steps that result in the production of the aryloxyl leaving group and a corresponding methoxylated

Scheme 1^a



^{*a*} $R = CH_3$ or 2-hydroxypropyl; -OR' = alkoxide.

phosphate.^{4,5} For both of these systems, the synergy created between a highly active dinuclear **3**:Zn(II)₂:($^{-}OCH_3$) catalyst and the methanol solvent accelerates the cleavage of phosphate diesters by 10^{11-13} times relative to the methoxide promoted background reactions at ${}^{s}_{s}$ PH 9.8 and 25 °C. That the rate enhancement far exceeds anything reported for this⁶ or related catalytic RNase or DNase models in water^{2,3,7} seems to be intimately tied to the reduced dielectric constant/polarity of the medium. Very recent work has shown that a different sort of medium effect provided by 80% DMSO/water confers very large rate accelerations of 2.7 × 10⁹ to 4.4 × 10¹⁰ for the Eu(III) and La(III) catalyzed hydrolysis of **1a** at essentially neutral pH in that medium.⁸

It seems possible that a further reduction of polarity/dielectric constant, while still retaining significant hydrogen bonding such as would be occasioned with ethanol, (dielectric constant $\varepsilon =$ 31.5 and 24.3 for methanol and ethanol respectively)⁹ might lead to even higher catalytic rate enhancements for the cleavage of phosphate diesters by 3:Zn(II)₂:(⁻OR).¹⁰ Herein we report a comprehensive study of the catalytic cleavage of diesters 1a-g promoted by $3:Zn(II)_2$ in anhydrous ethanol (99.9%). As will be seen, the kinetic pathways for all substrates proceed with a very strong substrate/catalyst saturation binding followed by a rate-limiting k_{cat}^{max} process which is shown to change from a chemical one for substrates with poor leaving groups, to a conformational change for those with good leaving groups. These are very fast reactions having k_{cat}^{max}/K_m terms 10^{12} to 10¹⁴ times larger than the corresponding second order rate constants for the ethoxide catalyzed reactions. Finally, in order to provide a deeper understanding of the catalytic process, we provide energetics calculations to determine the contributions of each of the kinetic and thermodynamic terms toward the acceleration achieved.

2. Experimental Section

2.1. Materials. Sodium ethoxide (21 wt % solution in denatured ethanol, titrated against N/50 Fisher Certified standard aqueous HCl solution and found to be 2.68 M) and Zn(CF₃SO₃)₂, were purchased from Aldrich and used without further purification. Tetrabutylammonium ethoxide in ethanol (~40%, titrated against 1 N Fisher certified standard aqueous HCl solution and found to be 1.08 \pm

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Table 1. Table of the Wavelengths Used to Monitor the **3**:Zn(II)₂:($^{\circ}$ OCH₃)-Catalyzed Reactions (λ_{cat}) and the Base Promoted Reactions (λ_{base}) of Phosphates **1** in Ethanol at 25 °C

		(•••••				-
phosphate	1a	1b	1c	1d	1e	1f	1g
$\lambda_{cat} (nm)$ $\lambda_{base} (nm)$	320 401	323 399	340 407	284 305	282	280	292

0.01 M) was obtained from Fluka. HClO₄ (70% aqueous solution, titrated to be 11.40 M) was purchased from Acros Organics and used as supplied. Anhydrous ethanol was purchased from Commercial Alcohols Inc. and was degassed by bubbling Ar through it for 1 h before storing it under Ar. The degassed ethanol was freshly dispensed prior to each set of kinetic experiments. Freshly dispensed ethanol was kept for a maximum duration of 1 h in an oven-dried, capped Erlenmeyer flask sealed with Parafilm between uses. The [H₂O] in the freshly dispensed degassed ethanol was found to be 0.028 ± 0.007 M using a Mettler Toledo DL32 Karl Fischer Coulometer, while the [H₂O] for the ethanol that has been kept in an Erlenmeyer flask for 1 h as described above (and used for experiments) was been determined to be 0.029 \pm 0.007 M. The sodium salts of aryl 2-hydroxylpropyl phosphates (1a-g) were prepared and characterized as described earlier.⁴ 1,3-Bis- N_1,N_1' -(1,5,9-triazacyclododecyl)propane (3) was prepared as described.^{5b} The dinuclear 3:Zn(II)₂:(-OCH₂CH₃) complexes were prepared as 2.5 mM stock solutions in degassed absolute ethanol by sequential addition of aliquots of stock solutions of sodium ethoxide, 1,3-bis- N_1, N_1' -(1,5,9-triazacyclododecyl)propane, and Zn(CF₃SO₃)₂ in stoichiometric ratios of 1:1:2. The complete formation of the active dizinc complex is achieved only after 50 min in ethanol (as monitored by the change in catalytic activity over time. This same phenomenon was observed for creation of the active forms of the diZn(II) and diCu(II) catalysts in methanol^{4,5}).

2.2. Methods. ¹H NMR and ³¹P NMR spectra were determined at 400 and 162.04 MHz. The CH₃OH₂⁺ and CH₃CH₂OH₂⁺ concentrations were determined using a combination glass electrode (Radiometer model XC100-111-120-161) calibrated with Fisher Certified standard aqueous buffers (pH = 4.00 and 10.00) as described in a previous paper.¹¹ ^s_spH values in ethanol were determined by subtracting a correction constant -2.54 from the readings obtained from the electrode, and the autoprotolysis constant of ethanol (K_{auto}) is taken to be $10^{-19.1.11}$

Literature ${}^{s}_{p}K_{a}$ values¹² of a series of different substituted phenols in ethanol and the measured half-neutralization ${}^{s}_{p}K_{a}$ values of *p*-nitrophenol and 2,4-dinitrophenol (0.5 mM of the phenols and 0.25 mM of NaOCH₂CH₃ in degassed absolute ethanol), were plotted against the aqueous pK_{a} values and found to fit a linear relationship, ${}^{s}_{p}K_{a}^{\text{EIOH}} = (1.24 \pm 0.01){}^{s}_{p}K_{a}^{\text{HOH}} + (3.2 \pm 0.1)$ (11 phenols; $r^{2} = 0.9990$) This relationship was used to interpolate the ${}^{s}_{p}K_{a}^{\text{EIOH}}$ values for the corresponding phenols of **1b,c,g** in ethanol.

2.3. Kinetics of Transesterification of 1a-g in Ethanol. The transesterification of phosphates 1a-g in degassed absolute ethanol were followed by following the rates of the appearance of the corresponding phenolic products by regular and stopped-flow UV/ visible spectrophotometry at 25.0 ± 0.1 °C at the wavelengths listed in Table 1.

For the complex-catalyzed reactions, a 2.5 mM stock solution of $3:Zn(II)_2:(\neg OCH_3CH_3)$ in degassed absolute ethanol was prepared in a capped and sealed (with Parafilm) oven-dried vial under N₂ at ambient temperature 1 h prior to the kinetic experiments to ensure the complete formation of the catalyst complex. This solution was used to prepare solutions of the catalyst with concentrations ranging from 0.02 mM $\leq [3:Zn(II)_2:(\neg OCH_2CH_3)] \leq 0.2$ mM, which were

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then loaded into one syringe of the stopped-flow reaction analyzer. Solutions of $(8-16) \times 10^{-5}$ M of substrates **1a**-g in ethanol were loaded into the second syringe. The final concentration of the phosphates was $(4-8) \times 10^{-5}$ M. At each [**3**:Zn(II)₂:($^{-}OCH_2CH_3$)] the pseudo-first-order rate constants (k_{obs}) were evaluated by fitting the UV/vis absorbance vs time traces to a standard exponential model. All reactions were followed to at least three half-times and found to exhibit good first-order rate behavior. At least five kinetic runs were conducted at each [**3**:Zn(II)₂:($^{-}OCH_2CH_3$)], with the reported k_{obs} values reported in Supporting Information being the average.

To determine the stability of the catalyst in the reaction medium, 8×10^{-5} M of **1a** in ethanol was loaded into one of the syringes in the stopped-flow reaction analyzer. An ethanol solution containing 0.8 mM of **3**:Zn(II)₂:($^{-}$ OCH₂CH₃) was loaded into the other syringe. Judging from the rate of appearance of the phenol product at 320 nm at different times over 200 min, there was no decomposition of the catalyst during this time.

During the above kinetic experiments, the ionic strength was not controlled nor was the spH controlled by buffers as we have found that the associated anions of these heavily inhibit the reactions. Thus, the spH values were set by the catalytic system itself and were generally found to be 9.0 ± 0.2 in the plateau region of the k_{obs} vs [3:Zn(II)₂:($^{-}OCH_2CH_3$)] plots. However, a $_{s}^{s}pH/log$ rate constant profile for the cleavage of 1e catalyzed by $3:Zn(II)_2$ in ethanol was conducted in the following manner. First, varying amounts of NaOCH2CH3 or HClO4 stock solutions (5 mM) in ethanol were added to solutions containing 2 \times 10⁻⁴ M of a preformed 3:Zn(II)₂:(⁻OCH₂CH₃) complex in ethanol which had been prepared an hour in advance to allow for complete formation of the catalyst complex. After the introduction of the additional acid or base the mixture was allowed to stand for 30 min to equilibrate (independent experiments showed that this time was optimum to attain the maximum kinetic activity) and then loaded into one of the two syringes of the stopped-flow analyzer while a 1.6×10^{-4} M solution of **1e** in ethanol was loaded into the other. The final concentrations after mixing in the reaction chamber were 1×10^{-4} M of **3**:Zn(II)₂ and 8×10^{-5} M of **1e** in ethanol. The ^s_spH values were measured at the end of the reactions. The spH/log rate constant profile plot given in Figure 5 shows a broad plateau from $_{s}^{s}$ pH 7.9–10, with the line through the data being derived from a fit of the solid squares data to a process that depended on two ionizable groups having ${}_{s}^{s}pK_{a}$ values of 7.2 and 10.8.

To determine the magnitude and type of $CF_3SO_3^-$ inhibition, a 4×10^{-5} M solution of 1a was premixed with varying concentrations of tetrabutylammonium triflate so that the final [triflate ion] ranged from $8 \times 10^{-4} - 4.8 \times 10^{-3}$ M, and the rate of the reaction in the presence of 0.2 mM of 3:Zn(II)₂:(⁻OCH₂CH₃) at each [triflate] was monitored in duplicate in ethanol, (^s_SpH = 8.54–8.85).

The rates of the ethoxide-catalyzed reactions of 1a-d ((1–2) × 10⁻⁴ M) were determined by UV/vis spectrophotometry at 25.0 ± 0.1 °C in the presence of tetrabutylammonium ethoxide at various concentrations between 0.004 and 0.4 M in degassed absolute ethanol. The kinetic data were analyzed by the initial rate method in which the first 2–10% of the Abs. vs time traces for appearance of phenolate products were fitted to a linear regression and the so-obtained rates converted to first-order rate constants (k_{obs}) by dividing them by the expected absorbance change if the reaction were to reach 100% completion. The reactions were carried out in duplicate and the plots of the average first-order rate constants (k_{obs}) vs [tetrabutylammonium hydroxide] were fitted to a standard linear regression model to find the second-order rate constants (k_{2} -OE).

3. Results

3.1. Ethoxide-Catalyzed Transesterification of 1a-d in Ethanol. The ethoxide promoted cleavages of **1a-d** in ethanol at 25.0 \pm 0.1 °C were studied under pseudo-first-order conditions of excess [CH₃CH₂O⁻] and the kinetics of formation of



Figure 1. Brønsted plot of log (k_2^{-OEt}) vs the ${}_{s}^{s}pK_{a}$ values for the 3:Zn(II)₂: (⁻OCH₂CH₃)-catalyzed cleavage of **1a**–**d** in degassed absolute ethanol at 25.0 ± 0.1 °C. The data fit a standard linear regression of $k_2^{-OEt} = (-0.90 \pm 0.04) {}_{s}^{s}pK_{a} + (7.2 \pm 0.6); r^2 = 0.9955.$

the phenolate products were monitored by initial rate methods due to the slowness of the reactions. Phosphates 1e-g react too slowly in base to determine the rate constants accurately in a reasonable time. In Figure 1 is a plot of the second order rate constants (k_2^{-OEt}) for substrates 1a-d vs the ${}_{s}^{s}pK_{a}$ of the corresponding phenols in ethanol (see Supporting Information). These data were fit by linear regression as $k_2^{-OEt} = (-0.90 \pm 0.04) {}_{s}^{s}pK_{a} + (7.2 \pm 0.6), r^2 = 0.9955; n = 4$, which was used to estimate the rate constants for 1e-g given in Table 2.

3.2. 3:Zn(II)₂:(⁻OCH₂CH₃)-Promoted Transesterification of 1 in Ethanol. Shown in Figure 2 is a representative plot of the uncorrected k_{obs} for reaction of **1a**-g vs total [**3:**Zn(II)₂: (⁻OCH₂CH₃)]_{*t*} put into the solution. The plots for all the substrates follow the same general appearance, in passing from substoichiometric amounts of catalyst to a roughly 2-fold excess of catalyst/substrate. All the plots show an apparent *x*-intercept and very strong 1:1 saturation binding superimposed on an inhibition curve that depends upon the increasing [⁻OTf] (each equivalent of catalyst brings with it 4 equiv of triflate).

We have previously demonstrated that triflate anion is a competitive inhibitor of the catalysis of phosphate diesters exhibited by **3**:Zn(II)₂:($^{-}OCH_3$) with a $K_i = 14.9$ mM in anhydrous methanol.^{5b} In ethanol, the affinity of triflate for the positively charged catalyst is enhanced and analysis of the k_{obs} vs [^{-}OTf] data shown in Figure 3 for the **3**:Zn(II)₂:($^{-}OCH_2CH_3$) promoted reaction of **1a** in ethanol gives an inhibition constant of $K_i = (0.36 \pm 0.02)$ mM. The analysis of the data is somewhat complicated and requires explanation to account for the appearance of the primary data plot in Figure 2 and the considerable inhibition provided by triflate.

The appearance of the triflate inhibition plot in Figure 3 suggests that this anion is an uncompetitive¹³ inhibitor following the simplified process given in Scheme 2 where K_i refers to the dissociation constant for the triflate inhibited complex, (**3**:Zn(II)₂: ($^{-}OCH_2CH_3$):**1**:(^{-}OTf)) in units of mM. The scheme is based on the assumption that the binding of the phosphate is far larger than that of triflate, and that there are two substrate-bound forms of the catalyst, namely (**3**:Zn(II)₂:($^{-}OCH_2CH_3$):**1**)_{free}, which leads to the product, and a triflate bound form, (**3**:Zn(II)₂:($^{-}OCH_2CH_3$):**1**)_{free}

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⁽¹⁴⁾ Since the triflate binding is quite strong, the more accurate method for determining the K_i is to use a variant of the universal binding equation (see Supporting Information). However, using this form lowers the values of the $k^{\text{obs corr.}}$ by only 4%, which is within the experimental uncertainty, so we have opted to use simplified method of eqs 1 and 2.

Table 2. Kinetic Data (Maximum Rate Constant for the $3:Zn(II)_2:(^{-}OCH_2CH_3)$ -Catalyzed Reactions Corrected for Triflate Inhibition ($k_{cat}^{max corr.}$), Second-Order Rate Constants for the Ethoxide-Promoted Reactions (k_2^{-OEt}), and the Catalytic Rate Accelerations ($k_{cat}^{max corr.}$ / K_M)/ k_2^{-OEt}) for the Cleavages of 1a-g in Ethanol at 25 ± 0.1 °C

phosphate diester	${}^{s}_{s}pK_{a}$ of phenol	$k_{\text{cat}}^{\text{max corr.}} (s^{-1})^a$	k_2^{-OEt} (M ⁻¹ s ⁻¹)	$(k_{\rm cat}^{\rm max\ corr.}/K_{\rm M})/k_2^{-{\rm OEt}c}$
1a	12.05	168 ± 6	$(2.43 \pm 0.09) \times 10^{-4}$	2.2×10^{12}
1b	12.39	133 ± 5	$(8.3 \pm 0.4) \times 10^{-5}$	5.1×10^{12}
1c	13.60	139 ± 3	$(9.7 \pm 0.7) \times 10^{-6}$	4.5×10^{13}
1d	14.83	36 ± 1	$(6.3 \pm 0.3) \times 10^{-7}$	1.8×10^{14}
1e	15.15	14.5 ± 0.3	3.7×10^{-7b}	1.2×10^{14}
1 f	15.60	4.5 ± 0.1	1.4×10^{-7b}	1.0×10^{14}
1g	15.92	2.67 ± 0.06	7.4×10^{-8b}	1.1×10^{14}

^{*a*} $k_{cat}^{max corr.}$ values are determined as described in text by fitting the k_{obs} vs [cat] data to eq 1 after correction for triflate inhibition ^{*b*} k_2^{-OEt} values for the base promoted reactions of the less reactive substrates 1e-g were estimated from the linear regression equation for the Brønsted plot in Figure 1. ^{*c*} Ratio of the apparent second order rate constant for the $3:Zn(II)_2:(-OCH_2CH_3)$ - promoted reaction of 1 (given as $k_{cat}^{max corr.}/K_m$) and second-order rate constant for the ethoxide reaction. K_m value represents the dissociation constant of $3:Zn(II)_2:(-OCH_2CH_3):1$ Michaelis complex which has an upper limit of $10^{-6.5}$ M. See text.



Figure 2. Plot of $k_{obs}^{(uncorr)}$ vs $[3:Zn(II)_2:(^{-}OCH_2CH_3)]_t$ for the catalyzed transesterification of 1g (5 × 10⁻⁵ M) at 292 nm and 25 °C in absolute ethanol.



Figure 3. Plot of k_{obs} vs [tetrabutylammonium triflate] for the catalyzed transesterification of **1a** (4 × 10⁻⁵ M) with [**3**:Zn(II)₂:($^{-}OCH_2CH_3$)]_t = 0.2 mM at 320 nm and 25 °C in degassed absolute ethanol. The inhibition constant (K_i) for triflate anion was determined by fitting the data into eq 1S, (see Supporting Information) and was found to be (0.36 ± 0.02) mM (dashed line represents the fit).

Scheme 2. Uncompetitive Triflate Anion ($^{-}$ OTf) Inhibition of the **3**:Zn(II)₂:($^{-}$ OCH₂CH₃)-Catalyzed Transesterification of 1 in Ethanol

3:Zn(II) ₂ :(⁻ OCH ₂ CH ₃)	$\underbrace{K_{m}}_{\text{(3:Zn(II)_2:(^{\circ}OCH_2CH_3):1)}_{free}}$	k _{cat}	Produc
+ 1	41		
	K _i OTf		
	" 3·Zn (II) ₂ ·("OCH ₂ CH ₂): 1 ·("OTf)		

1:⁻OTf), which has an insignificant reactivity relative to the latter. The first assumption seems justified by the fact that the $K_{\rm m}$ dissociation constants of all the (3:Zn(II)₂:(⁻OCH₂CH₃): **1a-g** complexes are very small and estimated to be at most 10^{-6.5} M *vide infra*. The second assumption is reasonable since the fit of the experimental data to eq 1S (see Supporting

Information) gives the line in Figure 3 that asymptotically approaches a limiting value of zero for the observed rate constant (within experimental uncertainty) for a fully triflate bound complex. The amount of free catalytically active species (3: Zn(II)₂:($^{-}OCH_{2}CH_{3}$):1)_{free} is calculated from eq 1 while eq 2 is used provide a rate constant for catalyzed cleavage of 1¹⁴ ($k_{obs}^{corr.}$) from the raw kinetic data ($k_{obs}^{uncorr.}$) after correction for the triflate inhibition.

$$[(\mathbf{3}: \operatorname{Zn}(\operatorname{II}):(\operatorname{OCH}_{2}\operatorname{CH}_{3}):\mathbf{1})]_{\text{free}} = \frac{(K_{i})[(\mathbf{3}: \operatorname{Zn}(\operatorname{II}):(\operatorname{OCH}_{2}\operatorname{CH}_{3}):\mathbf{1})]_{\text{total}}}{K_{i} + [\operatorname{OTf}]} \quad (1)$$

$$k_{\text{obs}}^{\text{corr.}} = \frac{k_{\text{obs}}^{\text{uncorr.}}(K_{i} + [\operatorname{OTf}])}{K_{i}} \quad (2)$$

Equation 3 is a universal expression¹⁵ applicable to both strong and weak binding situations that has been used to analyze kinetic data for similar systems in methanol.^{4,5} In eq 3, [sub] refers to the initial concentration of 1, [cat] refers to the concentration of viable catalyst, which is derived from the expression $[cat] = [cat]_{total} - A$, where A is an independently fitted parameter that corresponds to the x-intercept value observed in the kinetic plots of k_{obs} vs $[3:Zn(II)_2:(-OCH_2CH_3)]_t$ for all substrates 1. This intercept was previously observed in analogous plots obtained in methanol, and was explained by a dissociation of metal ion away from the catalyst at low concentrations which led to an inactive form.^{4,5} $K_{\rm B}$ is defined as the binding constant (units of M⁻¹) between the catalyst and 1. We define $K_{\rm m}$ (the reciprocal of $K_{\rm B}$) as the dissociation constant of the nontriflate-associated substrate:catalyst {(3: $Zn(II)_2:({}^{-}OCH_2CH_3):1)_{free}$ complex in units of M, with k_{cat}^{max} being the maximum rate constant going from the catalytically active species (3:Zn(II)2:(-OCH2CH3):1)free forward to the product.

$$k_{\rm obs} = k_{\rm cat} (1 + K_{\rm B} * [{\rm sub}] + [{\rm cat}] * K_{\rm B} - X) / [{\rm sub}] / (2K_{\rm B})$$
(3)

where:

$$X = \{(1 + 2K_{\rm B} * [{\rm sub}] + 2 * [{\rm cat}] * K_{\rm B} + K_{\rm B}^2 * [{\rm sub}]^2 - 2 * K_{\rm B}^2 * [{\rm cat}] [{\rm sub}] + [{\rm cat}]^2 * K_{\rm B}^2 \}^{0.5}$$

In Figure 4 is a plot of the $k_{obs}^{corr.}$ values vs [(3:Zn(II)₂: (⁻OCH₂CH₃)]_{total} for the catalyzed cleavage of **1g** in ethanol where, due to the strong binding, the plot breaks sharply at the point where there is a 1:1 ratio of catalyst (over the amount at the *x*-axis intercept, *A*) and phosphate. Similar plots are observed



Figure 4. Plot of $k_{obs}^{(corr)}$ vs [3:Zn(II)₂:($^{-}OCH_2CH_3$)] for the catalyzed cleavage of 1g (5 × 10⁻⁵ M) at 292 nm and 25 °C in absolute ethanol. The raw data (Figure 2) was corrected for triflate inhibition using eqs 1 and 2. By fitting the corrected data to eq 3, one gets the line through the data with the maximum rate constant (k_{obs}^{max} corr.) determined to be 2.67 ± 0.06 s⁻¹ and $A = (1.78 \pm 0.01) \times 10^{-5}$ M.

for the $3:Zn(II)_2:(\neg OCH_2CH_3)$ -catalyzed cleavage of each of 1a-g in ethanol (see Supporting Information) and the kinetic data are fitted to eq 3 to determine the $k_{cat}^{max corr.}$ values (after correcting for triflate inhibition): these are presented in Table 2.

Since the binding of the phosphate to the complex is so strong, accurate values for $K_{\rm B}$ cannot be obtained from the data at hand. However an upper limit can be estimated through an iterative procedure where the $K_{\rm B}$ value is increased until the goodness of fit maximized. For all substrates, the goodness of the fits did not change when the $K_{\rm B}$ value exceeded $10^{6.5}$ M⁻¹, so we have assumed an upper limit for the binding constant for all substrates of $K_{\rm B} = 3.2 \times 10^6$ M⁻¹ (correspondingly $K_{\rm m} = 3.2 \times 10^{-7}$ M).

A ^s_spH vs log $k_{obs}^{max corr.}$ profile (Figure 5) was constructed for the reaction of 1e promoted by $3:Zn(II)_2$ as follows. A 2 \times 10^{-4} M solution of the catalyst (3:Zn(II):($^{-}OCH_2CH_3$)) was prepared in ethanol, as described in the experimental section, and then treated with varying amounts of HClO₄ or NaOEt to vary the ^spH. The mixtures were allowed to equilibrate for 30 min and then mixed by stopped-flow with 1.6×10^{-4} M of phosphate 1e, and the kinetics of the reaction were monitored (final concentrations were half-those in the syringes). At these concentrations all the substrate is bound to catalyst so the observed rate constant corresponds to k_{obs}^{max} which is then numerically corrected for triflate inhibition. Following the reaction, the spH values of the mixtures were measured and assumed to be representative of those during the reaction. The $_{\rm spH/log}^{\rm s} k_{\rm obs}^{\rm max \ corr.}$ plot of Figure 5 shows a hint of bell-shape with a broad plateau between $\sim {}^{s}_{s} pH$ 7.9 and 10, and when the solid squares data are fit to a two ${}_{s}^{s}pK_{a}$ model (Scheme 3, eq 2S in Supporting Information), this gives a computed $k_{cat}^{max corr.}$ of 13.7 s⁻¹ (compare with the value of 14.5 ± 0.3 s⁻¹ in Table 2 determined in a different way) and ${}_{s}^{s}pK_{a}^{1}$ and ${}_{s}^{s}pK_{a}^{2}$ values of 7.2 and 10.8.¹⁶ Notable is the observation that below pH 7.7, the catalytic activity drops more precipitously than theory predicts, suggesting that the catalyst is not stable once the



Figure 5. ${}^{s}_{s}$ pH/log k_{obs}^{max} corr. profile for the reaction of **3**:Zn(II)₂: (⁻OCH₂CH₃) and **1**e conducted at 25 °C at respective concentrations of 1 $\times 10^{-4}$ M and 8 $\times 10^{-5}$ M. The line through the square data points is computed on the basis of the process given in Scheme 3 having a bell-shaped ${}^{s}_{s}$ pH/rate profile giving two fitted ${}^{s}_{s}$ pK_a values of 7.2 \pm 0.4 and 10.8 \pm 0.1. The shaded data below ${}^{s}_{s}$ PH 7.7 were not used for fitting.

removal of the ethoxide (or its kinetic equivalent) commences. Therefore, none of the data in the shaded area was used for any fits.

4. Discussion

4.1. Ethoxide-Promoted Reactions of Phosphates 1. The Figure 1 Brønsted plot for the ethoxide-promoted reactions of **1a–e** in ethanol has a β_{lg} value of -0.90 ± 0.04 which is slightly larger than the -0.72 ± 0.08^4 observed for the methoxide-promoted cyclization of 1a-g in methanol and the hydroxide-promoted cyclization of 1a g in methanol and the hydroxide-promoted cyclization of 2-hydroxypropyl aryl phos-phates in water (β_{lg} value of -0.62).¹⁷ There is considerable debate about whether the cleavage of phosphate diesters of type **1** is concerted or stepwise.^{18,19} However, a recent study of the OH-promoted cleavage of uridine 3'-phosphate esters in water suggested a stepwise cleavage mechanism with the rate-limiting step with good leaving groups being the cyclization step ($\beta_{lg} =$ -0.52) while that with poor leaving groups being the breakdown of the phosphorane ($\beta_{1g} = -1.34$).²⁰ In the latter report²⁰ it was suggested that the original data for the base promoted cyclization of 2-hydroxypropyl aryl and alkyl phosphates²¹ could be reinterpreted as being consistent with a stepwise process involving a five-membered cyclic phosphorane intermediate. The change in the rate-limiting step observed for such stepwise reactions occurs at the quasi-symmetrical point where the pK_a of the leaving group (HOR) is approximately the same as the pK_a of the nucleophilic 2-hydroxypropyl group. Since the ${}_{s}^{s}pK_a$ of the corresponding phenol leaving groups of phosphates 1a-d

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- (22) Sánchez-Lombardo and Yatsimirsky have estimated8 the *p*Ka of the 2-hydroxy group in **1a** as 14.9 in water. Based on the relationship ${}^{s}_{p}K_{a}^{EIOH} = (1.24 \pm 0.01)pK_{a}^{H2O} + (3.2 \pm 0.1)$ given in the Experimental Section herein, the computed ${}^{s}_{p}K_{a}^{EIOH}$ is 21.7.

⁽¹⁵⁾ Equation 3 was obtained from the equations for equilibrium binding and for conservation of mass by using the commercially available MAPLE software, Maple V Release 5; Waterloo Maple Inc.: Waterloo, Ontario, Canada, 2003.

⁽¹⁶⁾ The first ${}^{s}_{p}K_{a}$ is not well-defined by the data of Figure 5 due to the presumed decomposition of the catalyst. However, an experimentally similar ${}^{s}_{p}$ H vs log k_{obs} profile is seen for the cleavage of a series of aryl methyl phosphate esters in ethanol catalyzed by 3:Zn(II)₂:(OEt) ${}^{s}_{p}K_{a}{}^{1} = -7$ and ${}^{s}_{p}K_{a}{}^{2} = 10.8$; to be published.

are all lower than the ${}^{s}_{p}K_{a}$ of the 2-hydroxylpropyl group (estimated to be about 21.7 in ethanol)²² the k_2^{-OEt} constant for cyclization of all of **1a**-**g** should all fall on a Brønsted line corresponding to rate-limiting formation of a phosphorane intermediate. Assuming that the β_{eq} of -1.74 for the transfer of the phosphoryl group between oxyanions in water²³ can be extrapolated to ethanol, the Leffler parameter, $\alpha = \beta_{lg}/\beta_{eq} = 0.52$ for the ethoxide reactions of phosphates **1a**-**g** in ethanol, suggests that in the transition state for cyclization the P–OAr bonding character progress some 52% of the way from the starting material to phenolate product with the aryloxy oxygen now having a net charge of $\sim -0.16 = (+0.74-0.90)$ in the TS. Should the process really be concerted and proceed via a single TS, the analysis would be essentially the same suggesting that the TS is central, about halfway between starting material and product.

It is an expected consequence of reduced dielectric constant/ polarity that the rates of reactions between species of the same charge type are retarded, while those between oppositely charged species are accelerated. For the lyoxide-promoted reaction of 1a shown in eq 4, k_2^{-OEt} is $2.4 \times 10^{-4} \text{ M}^{-1}\text{s}^{-1}$, k_2^{-OMe} is $2.6 \times 10^{-3} \text{ M}^{-1}\text{s}^{-1.24}$ and that for the hydroxide promoted cyclization is reported to be $9.9 \times 10^{-2} \text{ M}^{-1}\text{s}^{-125}$ or $6.5 \times 10^{-2} \text{ M}^{-1}\text{s}^{-1.26}$ The 10-fold rate reduction in ethanol relative to methanol is a consequence of the reduced polarity/dielectric constant which opposes the pre-equilibrium formation of the dianionic form (1a⁻).



As will be shown later, this same reduced dielectric constant/ polarity effect considerably enhances the reactions of substrates 1 when they are bound to the positively charged $3:Zn(II)_2$: ($^{O}CH_2CH_3$) catalyst as $3:Zn(II)_2:(^{O}CH_2CH_3):1$ or the kinetic equivalent $3:Zn(II)_2:1^{-}$ in ethanol relative to methanol.

4.2. 3:Zn(II)₂:(⁻OCH₂CH₃)-Catalyzed Transesterification of **1.** The catalytically active form of the complex in this study is almost certainly stoichiometrically analogous to that found in methanol,^{4,5} comprising a 1:2:1 ratio of ligand:Zn(II)₂:(⁻OR). While **3:**Zn(II)₂:(⁻OCH₃) and **3:**Zn(II)₂:(⁻OCH₂CH₃) are stable when prepared as described in the Experimental Section, they form only slowly and so are not amenable to titrimetric studies to provide the thermodynamic ${}_{8}^{8}pK_{a}$ values. In past work we have estimated the first ${}_{8}^{8}pK_{a}$ by determining the ${}_{8}^{5}pH$ at half-neutralization immediately following the addition of half an

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Figure 6. Brønsted plot of log (k_{cat}^{max} corr.) vs the ${}^{s}_{p}K_{a}$ values of the corresponding aryl leaving groups for the **3**:Zn(II)₂:($-OCH_2CH_3$)-catalyzed cleavage of **1**a-g in absolute ethanol at 25 °C. The two lines cross at approximately ${}^{s}_{p}K_{a}$ = 14.3. The line was constructed by NLLSQ fitting all the data to an expression k_{cat}^{max} corr. = $k_1k_2/(k_{-1} + k_2) = C_1C_210^{(\beta 1+\beta 2)pK_a/}$, ($C_{-1}10^{\beta-1pK_a}+C_210^{\beta 2pK_a}$).²⁷ The two fitted β values are -1.12 ± 0.12 and 0.0 ± 0.1 .

Scheme 3. Proposed Process Depending on Two ${}^s_{s}pK_{a}$ Values for the Catalyzed Cleavage of 1 Promoted by Complex $3:Zn(II)_2:({}^{-}OCH_2CH_3)$

3:Zn(II) ₂ (EtOH) _n $\xrightarrow{K_a^+}$ 3:Zn(II) ₂ (⁻ OEt) + H ⁺ $\xrightarrow{K_a^2}$	3:Zn(II) ₂ (⁻ OEt) ₂
К _В И 1	$+ H^+$
$3:Zn(II)_{2}(\text{OEt}):1 \\ or \\ 3:Zn(II)_{2}:1^{-} \xrightarrow{k_{cat}} Pro$	oduct

equivalent of $HClO_4$ to a methanol solution of $3:Zn(II)_2$: (⁻OCH₃). However, in this study it was found that the partial ^spH/rate profile shown in Figure 5 could be obtained by adding small amounts of HClO₄ or NaOEt to a preformed catalyst, allowing this mixture to equilibrate for some time, and then determining the kobs max corr. for the kinetics of cleavage of substrate 1e which is fully bound to the complex as $3:Zn(II)_2$: $(^{-}OCH_2CH_3)$:1e or a possible kinetic equivalent, 3:Zn(II)₂:1e⁻. where the 2-hydroxy group is deprotonated. The spH of the mixtures were measured following determination of their kinetics and were assumed to be representative of that in the reacting solution. The log $k_{obs}^{max corr./s}$ pH data in Figure 5 follow an apparent bell-shaped profile with the catalytic activity plateauing between ^spH 7.9 and 10, suggesting that it is follows the process given in Scheme 3 with two ionizations having ${}_{s}^{s}pK_{a}$ values of \sim 7.2 and 10.8 determined from fitting the solid square data of the figure to an appropriate equation (see eq 2S, Supporting Information). The kinetic data (O) obtained in the low spH domain do not fit the theoretical model and indicate that once the ethoxide begins to be removed from the complex, the system is unstable, probably dissociating one of the Zn(II) ions with concomitant loss of activity. This is consistent with our experience that indicates one cannot form these complexes in methanol or ethanol without having one equivalent of alkoxide present along with the ligand prior to the addition of the metal ions.

4.2.1. Change in Rate-Limiting Step for the k_{cat}^{max} corr. Term. The Brønsted plot given in Figure 6 exhibits a sharp downward break in log k_{cat}^{max} corr. commencing at ${}_{sp}^{s}K_{a} \approx 14.3$ suggesting a change in rate-limiting step for the unimolecular term dealing with product formation from some form of catalyst: substrate complex. Unlike the case in methanol,⁴ all the kinetic

⁽²³⁾ The extent of breaking of the P–OAr bond in the TS can be measured by the Leffler parameter, α, which measures the change in the Brønsted β_{1g} for the TS relative to the β_{eq} for equilibrium transfers of the phosphoryl group between oxyanion nucleophiles. In the case of the transfer of the (RO)P(=O)O⁻ group **18a**, the β_{eq} value is-1.74 with the O–Ar oxygen in the starting material having a net effective charge of +0.74. For the cyclization reaction involving attack of the 2-hydroxypropyl oxyanion, the Leffler parameter, α is given as β_{1g}/ β_{eq} = 0.52 suggesting that the P–OAr cleavage is 52% of the way from starting material to product, this assuming that the β_{eq} determined in water can be transposed into ethanol.

data for the catalyzed cleavages of 1a-g in ethanol exhibit saturation behavior with very strong binding.

The fact that the k_{cat}^{max} corr. terms for substrates 1a-c containing good leaving groups are essentially independent of the ${}_{s}^{s}pK_{a}$ ($\beta_{lg} = \sim 0$) indicates that the process that limits the rate for those substrates cannot be dependent on any chemical step where changes in the bonding of the P–OAr linkage is prominent. This is consistent with a nonchemical step such as a rearrangement process (k_{2} in Scheme 1) becoming rate-limiting for substrates with good leaving groups (1a-c) while the rates of the reactions for substrates with poorer leaving groups (1d-g) are limited by some chemical step where there is a large dependence on the leaving group ($\beta_{lg} = -1.12$).

These data and those previously in methanol⁴ can be accommodated within the simplified model for the catalyzed reaction given in Scheme 1 with consideration of the effect of the reduced dielectric constant medium on the first equilibrium constant ($K_{-1} = k_{-1}/k_1$ in units of M). For the Debye-Hückel model for association of spherical ions in a medium of dielectric constant ε_r the electrostatic potential energy of interaction between oppositely charged ions is:

$$P.E. = (z_+e)(z_-e)/(4\pi\varepsilon_0\varepsilon_r r)$$
⁽⁵⁾

where *r* is the distance between the centers of the ions, z_+e and z_-e are their charges in coulombs (*e* is the proton charge), and ϵ_o is the dielectric constant of the solvent in question.²⁸ In passing from water, to methanol and then ethanol, each 1 kcal/ mol of potential energy of attraction increases by a factor of 2.5 and then 3.2, so there is a dramatic effect of reduced dielectric solvent on the binding of ions of opposite charge. That effect will increase the k_1 association rate constant and decrease the k_{-1} dissociation rate constant, consistent with the observed larger catalyst:substrate binding constant in ethanol relative to methanol. While the dielectric constant has an obvious role in increasing the binding, once the complex is formed the ensuing chemical transformation should be less sensitive to changes in ϵ_r since the reactants are in intimate contact with solvent being excluded.

The solvent effect on the binding steps can be numerically evaluated using an approach similar to what we used before to analyze the change in rate limiting step in the $3:Zn(II)_2$: ($^{O}CH_3$)-catalyzed cleavage of 1a-g in methanol.⁴ In that case, analysis of the kinetic data suggested that the change in rate limiting step resulted from the substrate dependent partitioning of the doubly activated phosphate complex (k_{cat}^{max} vs k_{-2}). Although the k_{cat}^{max} corr. values we determined here for catalyzed cleavage of substrates 1c-g with poorer leaving groups are very close to the analogous values determined in methanol, the observation that all substrates adhere to saturation kinetics in ethanol suggests there are important differences in the two

solvents. The application of Michaelis–Menten kinetics to the process of Scheme 1 where the first step is treated as an equilibrium gives:

$$\operatorname{rate} = \frac{\mathrm{d}P}{\mathrm{d}t} = \frac{k_{\operatorname{cat}}^{\max}[\mathbf{3}:\operatorname{Zn}(\operatorname{II})_2:(^{-}\operatorname{OCH}_2\operatorname{CH}_3)]_{\operatorname{free}}[\mathbf{1}]}{K_{\operatorname{m}} + [\mathbf{3}:\operatorname{Zn}(\operatorname{II})_2:(^{-}\operatorname{OCH}_2\operatorname{CH}_3)]_{\operatorname{free}}}$$
(6)

$$K_{\rm m} = \frac{k_{-1}}{k_1} \bullet \frac{k_{-2} + k_{\rm cat}^{\rm max}}{k_2} = K_{-1} \frac{k_{-2}}{k_2} + K_{-1} \frac{k_{\rm cat}^{\rm max}}{k_2}$$
(7)

The $K_{\rm m}$ term can be broken down into two components as defined in eq 7. With substrates 1d-g having poor leaving groups where k_{cat}^{max} is less than k_{-2} , K_m is approximated as $K_{-1}(k_{-2}/k_{-2})$ k_2) and has an average value of $\sim (9 \pm 1) \times 10^{-5}$ M in methanol and, since it is dominated by binding effects, is insensitive to the nature of the leaving group. In ethanol, the analogous data for all the substrates gives an upper limit of $K_{\rm m}$ of 3 × 10⁻⁷ M, which is a likely consequence of a reduction in the K_{-1} term pertaining to dissociation of phosphate away from 3:Zn(II)₂: (⁻OCH₂CH₃). The fact that the break in the plot of Figure 6 is at $k_{\rm cat}^{\rm max \ corr.} \approx 150 \ {\rm s}^{-1}$ suggests that this is the value for the proposed rearrangement step (k_2) to form the proposed doubly activated phosphate by binding to both Zn(II) ions. Using this value it can be calculated that $K_{-1}k_{-2}$ is $\sim 4.5 \times 10^{-5} \text{ M} \cdot \text{s}^{-1}$: assuming this same k_2 value obtains for the rearrangement step in methanol one calculates that $K_{-1}k_{-2}$ there would be 1.4 \times $10^{-2} \text{ M} \cdot \text{s}^{-1}$.

4.2.2. The ${}^{s}_{p}k_{a}$ Dependent $k_{cat}{}^{max \ corr.}$ Terms. The $k_{cat}{}^{max \ corr.}$ terms for the catalyzed cleavages of the bound substrates 1d-g adhere to a Brønsted relationship having a β_{1g} of -1.12 ± 0.12 which is experimentally the same as was found for the catalyzed cleavage of 1c-g in methanol $(-0.97 \pm 0.05)^{4}$ and perhaps slightly larger than for the ethoxide promoted cleavage of 1a-d in ethanol (-0.90 ± 0.04) . While the rate-limiting step is either a concerted displacement or formation of a phosphorane intermediate, a weak conclusion can be drawn that there is a slightly more extensive change in the P–OAr bond in the $3:Zn(II)_{2}:(-OCH_{2}CH_{3})$ -catalyzed cleavage relative to the ethoxide promoted cleavage.

Shown in Scheme 4 is a proposed mechanism slightly expanded from that presented in Scheme 1 for the catalyzed cleavage based on the information gained in ethanol. As a starting point we formulate the essential ethoxide in the $3:Zn(II)_2$ complex as bridging between the two metal ions based on the structural evidence gained with the $diZn(II)^4$ and $diCu(II)^{5c}$ in methanol. Due to the high binding constant between 1 and 3:Zn(II)₂:(⁻OCH₂CH₃), we propose formation of a large equilibrium amount of Comp 1 where there is binding of the phosphate to one of the metal ions with a loosening of the bridging ethoxide. This is followed by rearrangement step(s) where the phosphate becomes doubly activated by binding to both Zn ions. The k_2 step involves simultaneous removal of the coordinated ethoxide and deprotonation of the 2-hydoxypropyl group to yield Comp 2 which can undergo intramolecular cyclization (stepwise or concerted) to give the five-membered cyclic phosphate product.²⁹ An alternative rearrangement process via k_2' involves formation of Comp 2' having a coordinated ethoxide (bridging or singly coordinated), which subsequently acts as a general base to assist in the cyclization. A general base process is excluded in the case of a diZn(II) complex promoting cyclization of **1a** in water³⁰ but is proposed as the viable mechanism for the Ln(III) promoted cleavages of 1a in 80% DMSO/water,⁸ so it is possible that there is a shift in

⁽²⁷⁾ Neverov, A. A.; Sunderland, N. E.; Brown, R. S. Org. Biomol. Chem. 2005, 3, 65.

⁽²⁸⁾ Levine, I. N. *Physical Chemistry*, 4th ed.; McGraw-Hill, Inc.: New York, 1978; pp 276–281.

⁽²⁹⁾ The first formed phosphate product is the cyclic five-membered phosphate and the observed product is the phenol/phenoxide of the parent 1. We have observed that the cyclic phosphate opens up very rapidly (t_{1/2} ≈ 2 s, but at least 10-times slower than the reaction of the slowest of our substrates 1) in the presence of 3:Zn(II)₂ in methanol to form a kinetic mixture of 2-hydroxypropyl methyl phosphate and its isomeric (1-(hydroxymethyl)ethyl)methyl phosphate in a 30:70 mixture. While we have not checked the situation in ethanol, there is no reason to suspect that the cyclic phosphate will not react rapidly with ethanol in the presence of the catalyst. Tsang, W. Y; Edwards, D.; Melnychuk, S. A.; Liu, C.; Neverov, A. A.; Brown, R. S. manuscript in preparation.

Scheme 4. Proposed Mechanism for the Catalyzed Reaction in Ethanola



Liu et al.

^a Zn charges omitted for simplicity.

mechanism from specific catalysis to general catalysis brought about by the medium effects that give the fast reactions observed here and previously.^{4,5,8} Indeed the available evidence now allows us to rule out a specific base catalyzed process in ethanol where <u>external</u> ethoxide acts as the base to remove the proton from the 2-hydroxy group prior to cyclization. For the **3**:Zn(II)₂: ($^{-}OCH_2CH_3$)-catalyzed cleavage of **1e** at $^{s}_{s}PH$ 7.76, the observed $k_{cat}^{max \ corr.}$ is 10.6 s⁻¹. At that $^{s}_{s}PH$, the free [^{-}OEt] is 4.6 × 10⁻¹² M (K_{auto} of ethanol is 10^{-19.1}), ¹¹ so the rate constant for cleavage promoted by external ^{-}OEt would need to be 2.65 × 10¹² $M^{-1}s^{-1}$, exceeding the diffusion limit in ethanol³¹ by a factor of 265.

4.3. Energetic Considerations for the Catalysis. In Table 2 are presented $(k_{cat}^{max corr.} / K_m)/k_2^{-OEt}$ values indicating the catalytic reactions in ethanol are 10^{12} to 10^{14} larger than the corresponding ethoxide promoted reactions. Although the experimental k_{cat}^{max} values in ethanol and methanol for substrates **1c-g** are close to each other, the $(k_{cat}^{max}/K_m)/k_2^{-OMe}$ ratios for these substrates in methanol vary from about 4 \times 10 8 to 4 \times 10^{9.4} The kinetic data clearly indicate the apparent 10⁴ larger activity in ethanol stems from respective 10 to 100-fold and at least 100-fold reductions in both the k_2^{-OR} terms and K_m terms relative to their values in methanol. An alternative comparison that gives apparently spectacular accelerations relates the $k_{cat}^{max corr.}$ values to the presumed ethoxide reaction at the ^s_spH where the catalyzed reactions were conducted. The [-OEt] at ${}_{s}^{s}$ pH 9.0 is ~10⁻¹⁰ M, so that for **1c**-**g**, the catalytic acceleration would be 10¹⁷-fold!³² By this measure, the catalytic acceleration in ethanol is 10⁵ larger than in methanol, but this stems from three main factors, including the 10 to 100-times less reactivity of the alkoxide reaction in ethanol, the decreased autoprotolysis constant of ethanol relative to methanol $(10^{-16.77})$ and the lower working ^s_spH (9.0 vs 9.8).

The more thermodynamically correct method to evaluate the catalytic efficacy of enzyme- or synthetic catalyst-promoted

Scheme 5



reactions computes the free energy of binding of the catalyst to the transition state of the presumed lyoxide promoted reaction.^{3,33,34} Following the procedures we used to analyze the **3**:Zn(II)₂: (⁻OCH₃) promoted cleavages of **1a**-**g** and **2a**-**n** in methanol,^{4,5} we consider in Scheme 5 a cycle encompassing the **3**:Zn(II)₂: (⁻OCH₂CH₃) and ethoxide reactions for **1a**-**g** in ethanol: the definitions of the terms are given in ref 4. Equation 8³⁵ provides the calculated free energy of binding of the catalyst to the ethoxide:substrate complex ($\Delta\Delta G^{\dagger}_{stab}$), that is, [**3**:Zn(II)₂: (⁻OCH₂CH₃):**1**][‡] and [CH₃CH₂O⁻:**1**]^{‡,4,5}

$$\Delta\Delta G_{\text{stab}}^{\neq} = (\Delta G_{\text{Bind}} - \Delta G_{\text{m}} + \Delta G_{\text{cat}}^{\neq}) - \Delta G_{\text{Non}}^{\neq} = -RT \ln \left[\frac{(k_{\text{cat}}/K_{\text{m}})(_{\text{s}}^{\text{s}}K_{\text{a}}/K_{\text{auto}})}{k_{2}^{-\text{OEt}}} \right]$$
(8)

The $k_{cat}^{max \text{ corr.}}$ and $k_2^{-\text{OEt}}$ values are from Table 1 with the $k_2^{-\text{OEt}}$ values for the less reactive substrates **1e**-**g** being extrapolated from the Brønsted plot in Figure 1. The upper limit for the K_m values for all substrates was taken to be $10^{-6.5}$ M and the ${}_{\text{sp}}^{\text{sp}}K_a$ for the formation of **3**:Zn(II)₂:($^{-}\text{OCH}_2$ CH₃) from **3**:Zn(II)₂:(HOCH₂CH₃) was taken to be 7.2 from the fitting of the data shown in Figure 5. While there may be some error in the latter two numbers, this does not affect the general picture

⁽³⁰⁾ Yang, M.-Y.; Iranzo, O.; Richard, J. P.; Morrow, J. R. J. Am. Chem. Soc. 2005, 127, 1064.

⁽³¹⁾ Schwarz, H. A.; Gill, P. A. J. Phys. Chem. 1977, 81, 22.

⁽³²⁾ Since we know that the activity of 3:Zn(II)₂:(⁻OEt) is maintained down to §pH 7.9, the acceleration relative to the base promoted reaction at that §pH would be over 10¹⁸-fold.

⁽³³⁾ Wolfenden, R. Nature 1969, 223, 704.

⁽³⁴⁾ For applications of this to phosphate cleavage and other reactions see: Yatsimirsky, A. K. *Coord. Chem. Rev.* **2005**, *249*, 1997.

⁽³⁵⁾ Equation 8 is a correct form of the eq 5 in the original methanol manuscript⁴ where a typographical error appeared in the expression $(\Delta \Delta G_{\text{stab}}^{\pm} = (\Delta G_{\text{Bind}} + \Delta G_{\text{m}} + \Delta G_{\text{cal}}^{\pm}) - \Delta G_{\text{Non}}^{\pm})$ placing a (+)-sign in front of the ΔG_{M} term. Since K_{M} refers to the dissociation constant for the Michaelis complex, and we are interested in the binding energy of catalyst and 1, the correct form of the equation should be $-\Delta G_{\text{M}}$.

Table 3. Tabulation of the $(k_{cat}^{max}/K_m)({}^{s}_{b}K_a/K_{auto})$ Constants and the Computed Free Energies for the Formation of Catalytic Complexes $(\Delta G_{Bind} - \Delta G_M)$, the Free Energies of Activation for k_{cat}^{max} corr. (ΔG^{*}_{cat}) , and the Free Energies of Stabilization of the Ethoxide Transition State through Binding to **3**:Zn(II)₂ $(\Delta \Delta G^{*}_{stab})^{a}$ for the Catalyzed Reaction of Substrates **1a**-**g** at 25 °C in Ethanol

substrate	$(k_{cat}^{max \ corr.} \ /K_m) \ ({}^s_s K_a / K_{auto}) \ (M^{-2} s^{-1})^{b, c}$	$\Delta G_{ ext{Bind}} - \Delta G_{ ext{M}} ext{ (kcal/mol)}^d$	$\Delta {\it G}^{ m t}_{ m cat}$ (kcal/mol) e	$\Delta {\it G}^{*}_{ m Non}$ (kcal/mol) e	$\Delta\Delta G^{\ddagger}_{\mathrm{stab}}$ (kcal/mol)
1a	4.2×10^{20}	-25.1	14.4	22.3	-33.0
1b	3.3×10^{20}	-25.1	14.5	23.0	-33.6
1c	3.5×10^{20}	-25.1	14.5	24.2	-34.8
1d	9.0×10^{19}	-25.1	15.3	25.9	-36.5
1e	3.6×10^{19}	-25.1	15.8	26.2	-35.5
1f	1.1×10^{19}	-25.1	16.5	26.8	-35.4
1g	6.7×10^{18}	-25.1	16.8	27.1	-35.4

^{*a*} $\Delta\Delta G^{\ddagger}$ stab computed from application of kinetic and equilibrium constants to eq 8. ^{*b*} ($k_{cat}^{max \text{ corr.}}/K_m$)/ k_2^{-OEt} values from Table 1 where the K_m values for **1a-g** are assumed to have a upper limit of $10^{-6.5}$ M. ^{*c*} ${}_{8}^{*}K_a$ of $10^{-7.22}$ determined from the fit of the data in Figure 5 corresponding to the first ionization in Scheme 3; $K_{auto} = 10^{-19.1}$; ${}_{8}^{*}K_a/K_{auto} = 7.94 \times 10^{11}$ and corresponds to the binding constant of. –OEt and **3**:Zn(II)₂. ^{*d*} Computed as ($\Delta G_{Bind} - \Delta G_m$) = –*RT* ln((${}_{8}^{*}K_a/K_{auto})/K_m$). ^{*e*} Computed from $\Delta G^{\ddagger}_{cat} = -RT \ln(k_{cat}^{max \text{ corr.}}/(kT/h))$ or $\Delta G^{\ddagger}_{Non} = -RT \ln(k_2^{-OEt}/(kT/h))$ from the Eyring equation where (kT/h) = 6 × 10¹² s⁻¹ at 298 K.



Figure 7. Comparison of the activation energy diagram for the $3:Zn(II)_2:(^{-}OR)$ and RO^{-} -catalyzed cleavages of 1c in ethanol and methanol at standard state of 1 M and 25 °C showing the calculated energies of binding the alkoxide by $3:Zn(II)_2$, of binding of 1c to $3:Zn(II)_2:(^{-}OR)$ and the calculated activation energies associated with k_{cat}^{max} and k_2^{-OR} . Methanolysis data taken from ref 4.

greatly except for a small numerical uncertainty of the ΔG terms that depend on those constants. In Table 3 are given the $(k_{\text{cat}}^{\text{max corr.}}/K_{\text{m}})({}^{\text{s}}_{\text{s}}K_{\text{a}}/K_{\text{auto}})$ and the computed $\Delta\Delta G^{\ddagger}_{\text{stab}}$ values for the catalyzed reactions of $\mathbf{1a}-\mathbf{g}$.

The data in ethanol invite comparison with those determined earlier for the catalyzed reactions in methanol⁴ and shown in Figure 7 are the free energy data for the cleavage of 1c in both solvents at a standard state of 1 M and 25 °C. The most striking feature is that the $\Delta\Delta G^{\dagger}_{stab}$ in ethanol (ranging from -33 to -36.5 kcal/mol) is very large in absolute terms and some 11–13 kcal/mol more negative than for the same substrate in methanol. Figure 7 visually informs us that the main differences in the ΔG values in the two solvents arise from three terms: a much stronger binding in ethanol of alkoxide to **3**:Zn(II)₂ and of 1c to **3**:Zn(II):(⁻OR) as well as the ~65-fold slower reaction of alkoxide in ethanol than in methanol.

The first two terms place the three reaction components into a 10-11 kcal/mol deeper thermodynamic well when fully bound in ethanol, while the third raises the alkoxide reaction's transition state by about 2.5 kcal/mol. However, the surprising aspect is that the k_{cat}^{max} corr. term is about the same in both solvents, the ΔG^{\dagger}_{cat} being 14.5 and 14.3 kcal/mol. It is also very interesting that the free energy of the $(3:Zn(II)_2(^{-}OEt):1)^{\dagger}$ is lower than the ground-state for the uncomplexed reaction partners by 10-11 kcal/mol in ethanol while in methanol the corresponding TS is roughly isoenergetic with the free reaction partners.

It is worthwhile to attempt to compare the accelerations for the reactions achieved by this catalytic system in ethanol with what is achievable by phosphodiesterase enzymes, bearing in mind that the medium, substrates and, in particular, the leaving groups are different. The cleavage of 3',5'-UpA is reported to have a first order rate constant at pH 6, T = 25 °C of 5×10^{-9} s^{-1 36} while a rate constant of 2.2×10^{-11} s⁻¹ was observed for the cleavage of 3',5'-ApG moiety inside in a strand of deoxynucleotides at 23 °C and pH 6.³⁷ Since it is probable that

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both these processes are specific base catalyzed throughout the accessible pH regions,³⁸ respective second order rate constants for the base catalyzed process of 5×10^{-1} and 2.2×10^{-3} $M^{-1}s^{-1}$ are calculated which can be compared with the reported value of 2×10^{-3} $M^{-1}s^{-1}$ cleavage of UpU.^{39,40} Since enzymes that promote the cleavage of RNA type phosphodiesters typically have k_{cat}/K_M values of 10^6 to 10^8 $M^{-1}s^{-1}$,¹d the computed acceleration as measured by $(k_{cat} / K_M)/k_2^{-OH}$ for the enzyme catalyzed cleavage of RNA would be $\sim 10^7$ to 10^{11} . In this study, accelerations of $1-2 \times 10^{14}$ are seen for substrates 1d-g where the k_{cat}^{max} corr. term specifically refers to the chemical cleavage step of the bound substrate.

5. Conclusions

The catalytic acceleration for the cleavage of substrates **1** by a dinuclear Zn(II) catalyst in methanol and ethanol far exceeds anything so far reported for metal ion containing catalysts in aqueous solution.² It is notable that the reaction in question is not a hydrolytic process in any case, but rather an intramolecular cyclization so the importance of the solvent as a nucleophile is not relevant. It has been stated that the effective dielectric constants in enzyme active sites resemble those of organic solvents rather than water.^{41,42} In the present case, reductions in dielectric constant such as what happens when one proceeds from water, to methanol and then ethanol, seem to be a particularly effective strategy for accelerating the rate of metal catalyzed acyl and phosphoryl transfer reactions.⁴³ The dinuclear catalyst (**3**:Zn(II)₂) in water is reported⁶ to be poor, and in fact no more effective in promoting the hydrolysis of bis-pnitrophenyl phosphate than the Zn(II) complex of 1,5,9triazacyclododecane, so the strong activities seen in methanol and ethanol for phosphate diester cleavage^{4,5} point to a synergistic interaction between the catalyst and medium. It is an essential, but not exclusive, requirement that the catalyst must readily recruit the reaction partners $(3:Zn(II)_2 + 1 + OR)$ into a reactive complex and clearly the reduced polarity medium enhances these interactions between oppositely charged components. However, simple binding of the reaction partners cannot lead to rate accelerations unless there is a greater binding of the transition state for the catalyzed reaction. Indeed this binding amounts to 33-36 kcal/mol in while that in methanol is 21-23 kcal/mol, the difference in the two solvents being largely dependent on the far stronger binding of the anionic reactants by the positively charged dinuclear complex in ethanol. What is interesting is the fact that the k_{cat}^{max} terms are very similar in ethanol and methanol for the substrates where this term relates to a chemical cleavage step. Perhaps this results from the fact that once bound by the catalyst, the polar groups such as the metal ions and transforming phosphate are at the interior of the catalyst:substrate complex and so not subject to significant solvent effects.

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Supporting Information Available: Tables of pseudo-firstorder rate constants for reactions of 1a-g with $3:Zn(II)_2$: (OCH_2CH_3) or ethoxide in ethanol; plots of $k_{cat}^{max \text{ corr.}}$ vs the [$3:Zn(II)_2:(^{OCH_2CH_3})$] for the catalyzed reactions of 1a-g in ethanol, mathematical treatments for determining the triflate inhibition constants and for the fitting of the Figure 5 data to a two pK_a model. This material is available free of charge via the Internet at http://pubs.acs.org.

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(40) Substrates 1 have better leaving groups than the dinucleotides, but the computed background reactions for these at ^s₅PH9.0 are slower than that of the dinucleotides at pH 6 in water due to three main reasons. First the concentration of base at ^s₅PH 9 in ethanol is 8 × 10⁻¹¹ M while in water at pH 6 the [−OH] is 10⁻⁸ M⁻¹s⁻¹, as discussed in Section IV.1 due to electrostatic repulsion, the lyoxide reaction in ethanol is about 100 time slower than in water; third, model studies show that the 2-OH group in RNA provides about 10⁹ acceleration of the cleavage reaction while the acceleration provided in the 2 hydroxypropyl substrates is only about 10⁵. See ref 17.

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